Parasitenone, a New Epoxycyclohexenone Related to Gabosine from the Marine-Derived Fungus Aspergillus parasiticus

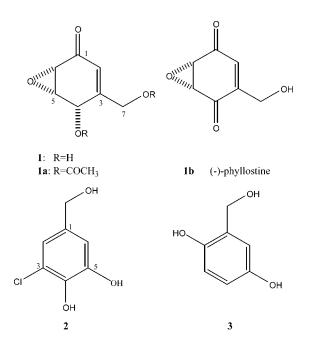
Byeng Wha Son,^{*,†} Jin Seok Choi,^{†,§} Jung Chul Kim,[†] Ki Wan Nam,[‡] Dong-Soo Kim,[§] Hae Young Chung,[⊥] Jung Sook Kang,[∥] and Hong Dae Choi[▽]

Department of Chemistry and Department of Marine Biology, Pukyong National University, Busan 608-737, Korea, Department of Food Science and Technology, Kyungsung University, Busan, Korea, College of Pharmacy and College of Dentistry, Pusan National University, Busan, Korea, and Department of Chemistry, Dongeui University, Busan, Korea

Received September 12, 2001

Bioassay-guided fractionation of an organic extract of the broth from the marine-derived fungus culture of Aspergillus parasiticus led to the isolation and subsequent structural elucidation of a new gabosine derivative, parasitenone (1), and two known benzyl alcohols, 3-chloro-4,5-dihydroxybenzyl alcohol (2) and gentisyl alcohol (3). The benzyl alcohols (2, 3) were identified as the principal free radical scavenging components. Parasitenone (1) also showed moderate activity in the free radical scavenging assay.

As part of an effort to discover biologically active natural products from marine organisms,¹ we investigated bioactive constituents of the marine algicolous fungus Aspergillus parasiticus and isolated a fairly unstable epoxycyclohexenone, parasitenone (1), and two known benzyl alcohols, 3-chloro-4,5-dihydroxybenzyl alcohol (2)² and gentisyl alcohol (3).³



Parasitenone (1) was isolated as a very unstable colorless oil, which was deduced to have the molecular formula $C_7H_8O_4$ from the HREIMS of its diacetate (1a) and ¹³C NMR data of 1 and 1a. The IR spectrum of 1 suggested the presence of hydroxyl (3356 cm^{-1}), enone (1680, 1027 cm⁻¹), and epoxy (1236, 903, 867 cm⁻¹) groups. The UV spectrum of **1** showed the presence of a $\beta\beta$ -disubstituted

10.1021/np010450k CCC: \$22.00

enone chromophore [203 nm (ϵ 5300), 237 (4800)]. Since 1a showed six degrees of unsaturation from HREIMS analysis, it was implied that 1a contains two rings. In the ¹H NMR spectrum, two proton signals [δ 5.79 (1H, d, J =6.2 Hz, 4-OH), 5.01 (1H, t, J = 5.5 Hz, 7-OH)] were exchanged by D₂O, suggesting that **1** had two hydroxyl groups. The ¹H and ¹³C NMR spectra of **1** show signals attributable to one ketone (δ 193.9, C-1), one trisubstituted double bond [(& 6.39, H-2), (& 141.4, C-2), (& 133.8, C-3)], one oxygenated methylene [(δ 4.07, 3.96, H₂-7), (δ 57.3, C-7)], and three oxygenated methines [(δ 4.70, H-4), (δ 63.7, C-4), (& 3.40, H-5), (& 52.9, C-5), (& 3.76, H-6), (& 54.0, C-6)] bearing oxygen. The upfield oxygenated ¹³C NMR signals (δ 52.9, C-5; δ 54.0, C-6) were assigned to an epoxy group.

Interpretation of 2D NMR (COSY, HMQC, HMBC) confirmed the presence of functional groups noted above and led to structure 1.

The physicochemical data of 1 were not consistent with the data for the 4-epimer of 1, which was previously reported as a synthetic intermediate in the synthesis of (–)-phyllostine (1b).⁴ So, we elucidated the stereostructure of 1.

The relative stereochemistry of 1 was determined by coupling constants ($J_{4,5} = 0$, $J_{5,6} = 4.2$ Hz, $J_{4,6} = 2.8$ Hz) and NOESY data. Key NOE correlations from H-4 to H-6 and from H-5 to H-6 indicated that H-4, H-5, and H-6 were all of the cis configuration. The three stereocenters in parasitenone (1) were assigned S configurations by interpretation of its CD data. Parasitenone (1) showed a positive first Cotton effect at 338 nm ($\Delta \epsilon + 0.95$), which from the octant rule indicated 4S, 5S, and 6S configurations for 1.5 The absolute configuration of **1** was further supported by chemospecific oxidation of the secondary allylic hydroxyl of 1 with pyridinium dichromate (PDC) in DMF to 1b, which was identified by direct comparison of its spectral data with those of natural (–)-phyllostine (1b).^{4,6} On the basis of the above evidence, the structure of parasitenone was determined to be (4S,5S,6S)-5,6-epoxy-4-hydroxy-3hydroxymethylcyclohex-2-en-1-one (1).

The polyoxygenated cyclohexenone derivatives known as gabosines have been previously reported from natural sources.^{7,8} Since they are biologically interesting for their phytotoxic, antibiotic, antitumor, and antigermination activities, they have been used as targets or intermediates for the synthesis of biologically active compounds.^{8,9}

© 2002 American Chemical Society and American Society of Pharmacognosy Published on Web 05/03/2002

^{*} To whom correspondence should be addressed. Tel: +82-51-620-6378. * Io whom correspondence should be addressed. 1el: +82-51-62
 Fax: +82-51-628-8147. E-mail: sonbw@pknu.ac.kr.
 † Department of Chemistry, Pukyong National University.
 * Department of Marine Biology, Pukyong National University.
 * Kyungsung University.
 L College of Pharmacy, Pusan National University.
 * College of Dentistry, Pusan National University.
 * College of Dentistry, Pusan National University.

Dongeui University.

There is considerable recent evidence that free radicals induce oxidative damage to biomolecules. This oxidative damage is considered to play a causative role in aging and several degenerative diseases such as Alzheimer's disease, rheumatoid arthritis, cancer, and atherosclerosis.¹⁰ So far, a number of free radical scavengers have been found by using various screening systems.^{10–12}

Parasitenone (1) and benzyl alcohols (2, 3) showed scavenging activities toward free radicals, 1,1-diphenyl-2picrylhydrazyl (DPPH), peroxy nitrite (ONOO⁻), superoxide radical $(^{\circ}O_{2}^{-})$, and nitric oxide radical $(^{\circ}NO)$. Parasitenone (1) exhibited mild activity against DPPH and ONOO⁻, with IC₅₀ values of 57.0 and 52.6 μ M, respectively. In contrast, 2 and 3 showed strong activity, and the corresponding IC₅₀ values (μM) for each radical are as follow: **2**, DPPH (0.6), ONOO⁻ (3.1), •O₂⁻ (11.0), •NO (0.5); **3**, DPPH (1.4), ONOO⁻ (2.2), ${}^{\bullet}O_2^-$ (50«, inactive), ${}^{\bullet}NO$ (0.4). Among the assay results obtained, gentisyl alcohol (3) exhibited more potent activity (300-fold) than the positive control (carboxy-PTIO), with an IC₅₀ value of 137.7 μ M against nitric oxide radical (NO). Details of the radical scavenging activity of compounds 1-3 and its mode of action are currently under investigation and will be reported in due course.

Experimental Section

General Experimental Procedures. Melting points were determined on a Electrothermal model IA 9100 micro-melting point apparatus and are uncorrected. Optical rotation was determined on a Perkin-Elmer model 341 polarimeter. The IR spectrum was recorded on a Bruker FT-IR model IFS-88 spectrometer. ¹H (400 MHz) and ¹³C NMR (100 MHz) spectra were obtained on a JEOL JNM-ECP 400 NMR spectrometer, using TMS or solvent peaks as reference standard. MS spectra were obtained on a JEOL JMS-700 spectrometer. UV/visible spectra were measured on a Hitachi U-2001 UV/vis spectrometer. CD spectra were taken on a JASCO J-715 spectropolarimeter.

Fungal Isolation and Culture. The fungal strain (culture # MFA 153) was isolated from the surface of the marine red alga Carpopeltis cornea collected in the Golmae village, Ulsan City, in 1999 and identified as a Aspergillus parasiticus on the basis of fatty acid methyl ester analysis (Korean Culture Center of Microorganisms, Seoul, Korea), similarity index 0.858. The fungus was cultured (20 L) for 30 days (static) at 29 °C in SWS medium: soytone (0.1%), soluble starch (1.0%), agar (1.5%), and seawater (100%).

Isolation of Parasitenone (1) and Benzyl Alcohols (2, 3). The resulting broth and mycelium were separately extracted with EtOAc and CH₂Cl₂-MeOH (1:1) to afford crude extracts of 0.6 and 2.2 g, respectively. The broth extract showed strong radical scavenging activity against DPPH, and the active components were purified by assay-guided isolation using repeated silica gel (n-hexane-EtOAc) and HPLC (10:1 EtOAc-MeOH) methods to yield a new gabosine derivative, parasitenone (1, 8 mg), as well as the known benzyl alcohols, 3-chloro-4,5-dihydroxybenzyl alcohol (2, 22 mg) and gentisyl alcohol (3, 12 mg).

Parasitenone (1): unstable colorless oil; $[\alpha]_D + 71.6^\circ$ (*c* 0.3, MeOH); IR (neat) v_{max} 3356, 1680, 1400, 1236, 1027, 903, 867 cm^-1; UV (MeOH) $\lambda_{\rm max}$ (log $\epsilon) 203$ (3.72), 237 (3.68) nm; CD (MeOH) ($\Delta \epsilon$) 338 (+0.95), 245 (-1.76) nm; CIMS *m*/*z* 156 [M]⁺ (100), 138 $[M - H_2O]^+$ (7), 122 $[M - H_2O - O]^+$ (2), 110(3), 86(3); ¹H NMR (400 MHz, DMSO- d_6) δ 6.39 (1H, dddd, J = 2.7, 2.6, 2.2, 2.1 Hz, H-2), 4.70 (1H, ddddd, J = 6.2, 2.8, 2.8, 2.6, 2.6 Hz, H-4), 5.79 (1H, d, J = 6.2 Hz, 4-OH), 3.40 (1H, d, J = 4.2 Hz, H-5), 3.76 (1H, ddd, J = 4.2, 2.8, 2.7 Hz, H-6), 3.96 (1H, dddd, J = 15.2, 5.5, 2.8, 2.2 Hz, H-7), 4.07 (1H, dddd, J = 15.2, 5.5, 2.8, 2.2 Hz, H-7)*J* = 15.2, 5.5, 2.6, 2.1 Hz, H-7), 5.01 (1H, t, *J* = 5.5 Hz, 7-OH); ¹³C NMR (100 MHz, DMSO-*d*₆) δ 193.9 (s, C-1), 141.4 (d, C-2), 133.8 (s, C-3), 63.7 (d, C-4), 52.9 (d, C-5), 54.0 (d, C-6). 57.3 (t, C-7).

Acetylation of Parasitenone (1). Acetylation of 1 (10 mg) in the usual manner (acetic anhydride-pyridine) furnished the diacetate (1a) (6 mg).

1a: colorless oil; IR (neat) ν_{max} 1737, 1367, 1208, 1023 cm⁻¹; ¹H NMR (400 MHz, DMSO- d_6) δ 6.49 (1H, dddd, J = 2.5, 2.2,1.5, 1.5 Hz, H-2), 5.90 (1H, dddd, J = 2.5, 2.2, 1.5, 1.5 Hz, H-4), 3.58 (1H, d, J = 4.2 Hz, H-5), 3.93 (1H, ddd, J = 4.2, 2.5, 2.5 Hz, H-6), 4.65, 4.59 (each 1H, ddd, J = 13.5, 1.5, 1.5 Hz, H2-7), 2.15, 2.03 (each 3H, s, 4- and 7-OAc); ¹³C NMR (100 MHz, DMSO-d₆) & 192.1 (s, C-1), 139.6 (d, C-2), 130.9 (s, C-3), 66.5 (d, C-4), 51.6 (d, C-5), 54.9 (d, C-6), 59.9 (t, C-7), 169.9, 169.8 (each s, 4- and 7-OAc), 20.8, 20.7 (each q, 4- and 7-OAc); EIMS *m*/*z* 240 [M]⁺ (5), 180 [M – AcOH]⁺ (15), 138 [M – AcOH $CH_2=C=O^{+}$ (100); HREIMS m/z 240.0641 (calcd for $C_{11}H_{12}O_6$, 240.0634).

Oxidation of Parasitenone (1). Pyridinium dichromate (67 mg, 0.17 mmol) was added to a solution of 1 (13 mg, 0.08 mmol) in DMF (1 mL) at 0 °C, and the mixture was stirred for 3 h at the same temperature, then at room temperature for 3 h. The reaction mixture was filtered through a Celite pad. The filtrate was reduced under vacuum, and the residue was chromatographed on a Si gel column eluted with n-hexanes-EtOAc (1:1) to afford (-)-phyllostine (1b) (2.7 mg).

(-)-Phyllostine (1b): reddish solid. The compound exhibits spectral data (mp, $[\alpha]_D$, IR, ¹H NMR) comparable to published values.4,6

Acknowledgment. NMR and mass spectral data were kindly provided by the Korea Basic Science Institute, Taejeon, and The Cooperative Laboratory Center of Pukyong National University. This study was supported by a grant of the Korea Health 21 R&D Project, Ministry of Health & Welfare, Republic of Korea (HMP-00-B-21600-0121).

Supporting Information Available: The physicochemical data of 3-chloro-4,5-dihydroxybenzyl alcohol (2) and methods of the radical scavenging assay. This material is available free of charge via the Internet at http://pubs.acs.org.

References and Notes

- Son, B. W.; Kim, J. C.; Choi, H. D. *Lipids* **2001**, *36*, 427–429.
 Neilson, A. H.; Allard, A.-S.; Hynning, P.-A.; Remberger, M. Appl.
- Environ. Microbiol. 1988, 54, 2226-2236.
- Sakamura, S.; Chida, T.; Ito, J.; Sakai, R. Agric. Biol. Chem. 1971, 35, 1810-1811. (3)
- Yoshida, N.; Konno, H.; Kamikubo, T.; Takahashi, M.; Ogasawara, K. Tetrahedron: Asymmetry 1999, 10, 3849-3857.
- (5) Imahori, K. Optical Activity-The Theory and Application, Tokyo Kagaku Dojin: Tokyo, 1979; pp 67–76.
- Sakāmura, S.; Ito, J.; Sakai, R. Agric. Biol. Chem. 1971, 35, 105-(6)110.
- (7)Bach, G.; Breiding-Mack, S.; Grabley, S.; Hammann, P.; Hütter, K.; Thiericke, R.; Uhr, H.; Wink, J.; Zeeck, A. Liebigs Ann. Chem. 1993, 241-250.
- (8) Kamikubo, T.; Hiroya, K.; Ogasawara, K. Tetrahedron Lett. 1996, 37, 499-502, and references therein.
- (9) Lubineau, A.; Billault, I. J. Org. Chem. **1998**, 63, 5668–5671.
 (10) Chung, H. Y.; Choi, H. R.; Park, H. J.; Choi, J. S.; Choi, W. C. J.
- Agric. Food Chem. 2001, 49, 3614-3621, and references therein. (11) Pietta, P.-G. J. Nat. Prod. 2000, 63, 1035-1042.
- (12) Hwang, B. Y.; Kim, H. S.; Lee, J. H.; Hong, Y. S.; Ro, J. S.; Lee, K. S.; Lee, J. J. J. Nat. Prod. 2001, 64, 82–84, and references therein.

NP010450K